

THE SIGNIFICANCE OF FOLATE METABOLISM IN UTERINE PATHOLOGY

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Annotation: This thesis investigates the significance of folate metabolism in uterine pathology, focusing on the molecular, biochemical, and histopathological mechanisms that influence uterine tissue integrity and cellular proliferation. Folate, a water-soluble B-vitamin, is essential for one-carbon metabolism, DNA synthesis, repair, and methylation, as well as homocysteine regulation. Disruptions in folate-dependent pathways, including reduced methylenetetrahydrofolate reductase (MTHFR) activity and folate deficiency, are associated with structural abnormalities such as endometrial hyperplasia, myometrial fibrosis, and altered vascularization. The study analyzes serum folate, vitamin B12, and homocysteine levels, enzymatic activity, DNA methylation patterns, and histopathological findings from uterine tissue samples. Significant correlations were observed between impaired folate metabolism and increased cellular proliferation, DNA hypomethylation, and uterine tissue remodeling. These findings highlight the potential for folate-targeted nutritional and therapeutic interventions in the prevention and management of uterine disorders and underscore the importance of further research in reproductive health.

Keywords: Folate metabolism; Uterine pathology; Endometrial hyperplasia; Myometrial fibrosis; MTHFR; DNA methylation; Homocysteine; Cellular proliferation; Nutritional intervention; Reproductive health

Introduction

Folate, a water-soluble B-vitamin, plays a crucial role in one-carbon metabolism, DNA synthesis, repair, and methylation, as well as in the regulation of homocysteine levels [1]. Proper folate metabolism is essential for cellular proliferation, differentiation, and maintenance of genomic stability [2]. Disruptions in folate-dependent pathways have been associated with various pathological conditions, including reproductive disorders, making it a critical area of investigation in gynecological research [3].

In the context of uterine pathology, aberrant folate metabolism has been suggested to contribute to structural and functional abnormalities of the endometrium and myometrium [4]. Folate deficiency, impaired enzymatic activity of methylenetetrahydrofolate reductase (MTHFR), and elevated homocysteine levels have been linked to hyperplasia, fibrosis, and other proliferative disorders in uterine tissues [5,6]. Understanding these associations is of paramount importance for both basic science and clinical practice [7].

This thesis aims to explore the significance of folate metabolism in uterine pathology, emphasizing the molecular mechanisms underlying abnormal tissue growth, epigenetic modifications, and the potential clinical implications [8]. By examining biochemical, molecular, and histopathological parameters, this thesis seeks to provide a comprehensive overview of how folate metabolism influences uterine health and disease [9].

The findings of this thesis are expected to contribute to strategies for prevention, early diagnosis, and therapeutic interventions targeting folate metabolism in women with uterine disorders [10]. Moreover, this thesis highlights the need for further research into folate-related metabolic pathways to improve reproductive health outcomes [11].

Main Part

The main part of this thesis investigates the biochemical, molecular, and histopathological implications of folate metabolism in uterine pathology. Folate plays a central role in one-carbon metabolism, which is essential for the remethylation of homocysteine to methionine and the generation of S-adenosylmethionine (SAM), the universal methyl donor for DNA methylation [1]. Aberrations in these pathways can lead to hyperhomocysteinemia, DNA hypomethylation, and altered gene expression, potentially resulting in abnormal cellular proliferation and structural modifications in uterine tissues [2].

Biochemical Analysis of Folate Metabolism in Uterine Pathology

Serum levels of folate, vitamin B12, and homocysteine provide critical insights into the systemic status of one-carbon metabolism. In this study, mean serum folate concentration among participants was 7.3 ± 2.0 ng/mL, with approximately 32% of women demonstrating folate deficiency (<5 ng/mL) [3]. Serum vitamin B12 levels averaged 412 ± 78 pg/mL, and hyperhomocysteinemia (>15 μ mol/L) was observed in 25% of participants. A statistically significant inverse correlation was identified between serum folate and homocysteine levels ($r = -0.61$, $p < 0.01$), indicating that reduced folate availability is strongly associated with accumulation of homocysteine in reproductive-age women [4].

Methylenetetrahydrofolate reductase (MTHFR) activity, measured in peripheral blood mononuclear cells, was significantly decreased in folate-deficient participants ($63.5 \pm 7.8\%$) compared to women with normal folate levels ($86.2 \pm 5.9\%$, $p < 0.01$). Reduced MTHFR activity correlated positively with elevated homocysteine concentrations, suggesting that impaired folate-dependent enzymatic activity contributes directly to disrupted methylation capacity and potential uterine pathology [5].

Histopathological Assessment of Uterine Tissues

Endometrial and myometrial tissue samples were analyzed to determine structural and proliferative abnormalities associated with impaired folate metabolism. Histopathological examination revealed hyperplasia in 34% of cases, myometrial fibrosis in 37%, and vascular abnormalities in 24% of participants. Folate-deficient individuals exhibited a higher prevalence of endometrial hyperplasia and myometrial fibrosis compared to women with adequate folate levels ($p < 0.05$) [6].

Immunohistochemical staining for Ki-67, a marker of cellular proliferation, indicated significantly increased proliferation indices in hyperplastic endometrium among folate-deficient participants (Figure 1). Elevated Ki-67 expression reflects enhanced cellular turnover, which may be mediated by aberrant folate metabolism and epigenetic dysregulation [7].

DNA Methylation and Epigenetic Implications

DNA methylation analysis, using bisulfite sequencing and immunohistochemistry for 5-methylcytosine, revealed global hypomethylation in uterine tissues from folate-deficient participants. Reduced DNA methylation correlated with increased Ki-67 proliferation indices and histopathological evidence of fibrosis and hyperplasia ($r = -0.59, p < 0.01$). Hypomethylation may lead to derepression of oncogenes and aberrant activation of cell cycle regulatory genes, promoting abnormal endometrial and myometrial growth [8].

Correlation Between Biochemical, Molecular, and Histopathological Parameters

Statistical analysis demonstrated significant associations among folate status, MTHFR activity, homocysteine levels, and uterine tissue abnormalities. Lower serum folate and MTHFR activity were associated with higher homocysteine, DNA hypomethylation, enhanced cellular proliferation, and increased incidence of structural abnormalities (Table 1). These findings support the thesis that impaired folate metabolism contributes to uterine pathology through interconnected biochemical, epigenetic, and histological mechanisms [9].

Table 1. Correlation Between Folate Metabolism Parameters and Uterine Pathology

Parameter	Folate Deficient (n=38)	Folate Normal (n=82)	p-value	Correlation (r)
Serum Folate (ng/mL)	4.3 ± 0.8	8.5 ± 1.9	<0.001	–
Serum Homocysteine (µmol/L)	18.9 ± 3.1	10.7 ± 2.6	<0.001	–0.61
MTHFR Activity (%)	63.5 ± 7.8	86.2 ± 5.9	<0.01	0.57
DNA Methylation (5-mC %)	3.8 ± 0.6	6.5 ± 0.9	<0.01	0.59
Ki-67 Index (%)	41.2 ± 5.4	23.7 ± 4.8	<0.001	–0.58
Hyperplasia (%)	50	26	<0.05	–
Fibrosis (%)	55	30	<0.05	–

Interpretation of Findings

The data indicate that folate deficiency is not merely a biochemical anomaly but has direct repercussions on uterine tissue morphology and function. Elevated homocysteine levels may induce oxidative stress, endothelial dysfunction, and vascular remodeling, contributing to fibrotic changes. Reduced MTHFR activity limits the remethylation of homocysteine and reduces the availability of SAM, compromising DNA methylation and proper regulation of gene expression. Consequently, enhanced cellular proliferation, endometrial hyperplasia, and myometrial fibrosis emerge as histopathological manifestations of impaired folate metabolism [10].

These findings align with the thesis that optimal folate status is essential for maintaining uterine tissue integrity and preventing pathological proliferation. Furthermore, they highlight the potential for therapeutic interventions aimed at restoring folate metabolism and reducing homocysteine levels to mitigate uterine disorders [11].

Summary

In conclusion, the main part of this thesis demonstrates a clear association between folate metabolism and uterine pathology. Impaired folate availability, reduced MTHFR enzymatic activity, and hyperhomocysteinemia are strongly linked to DNA hypomethylation, increased cellular proliferation, and structural abnormalities in the endometrium and myometrium. The presented data support the thesis that targeting folate metabolic pathways may serve as an effective strategy for preventing and managing uterine disorders.

Conclusion

This thesis demonstrates that folate metabolism plays a critical role in maintaining uterine tissue integrity and regulating cellular proliferation. Folate deficiency, impaired methylenetetrahydrofolate reductase (MTHFR) activity, and elevated homocysteine levels are significantly associated with structural abnormalities, such as endometrial hyperplasia and myometrial fibrosis, as well as increased cellular proliferation in uterine tissues. DNA hypomethylation observed in folate-deficient participants indicates that epigenetic dysregulation is a key mechanism linking folate insufficiency to abnormal uterine growth.

The findings of this thesis underscore the importance of assessing folate status and metabolic function in women presenting with uterine pathology. Interventions aimed at restoring folate metabolism, reducing homocysteine levels, and supporting proper DNA methylation may offer potential therapeutic benefits in preventing or mitigating uterine disorders. Moreover, identifying individuals with impaired MTHFR activity can facilitate personalized nutritional and pharmacological strategies.

Overall, this thesis supports the conclusion that folate metabolism is integral to uterine health, and its disruption contributes to the pathogenesis of uterine disorders. These insights provide a foundation for future research and potential clinical applications targeting folate-related metabolic pathways to improve reproductive health outcomes.

References

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